

Potential of (portable) NMR and MRI for plant phenotyping

Henk Van As



NMR/MRI and plant phenotyping

- Non-invasive NMR and MRI sensors to measure plant performance and uncover genetic determinants of enhanced agronomic traits.
- A number of approaches and parameters are available from such sensors to study complex plant physiological traits such as growth, biomass accumulation and yield throughout the plant (crop) growing season.
- Both below-ground (3D root architecture), above-ground (stem, leaves, water status, growth, fruit development) and transport traits can be studied.

Suggested NMR/MRI accessible traits

Table 1. Conceptual overview showing how better understanding or quantification of specific below-ground, above-ground or transport traits will can contribute to improve light-use-efficiency (LUE), water-use-efficiency (WUE) or nitrogen-use-efficiency (NUE)

	Phenotype	Method/approach	Improvement of
Above ground	Energy conversion in light reaction	Fluorescence, imaging spectroscopy	LUE
	3-D canopy structure	Stereo imaging	LUE
Transport/allocation	Sap flow	MRI, velocity radiotracers	WUE, NUE
	Plant water status	NMR	WUE
Below ground	Root architecture	2-D rhizotrons, 3-D MRI	NUE, WUE

Rascher et al, 2011, Functional Plant Biol. 38

There are some more possibilities

(TD-)NMR and MRI

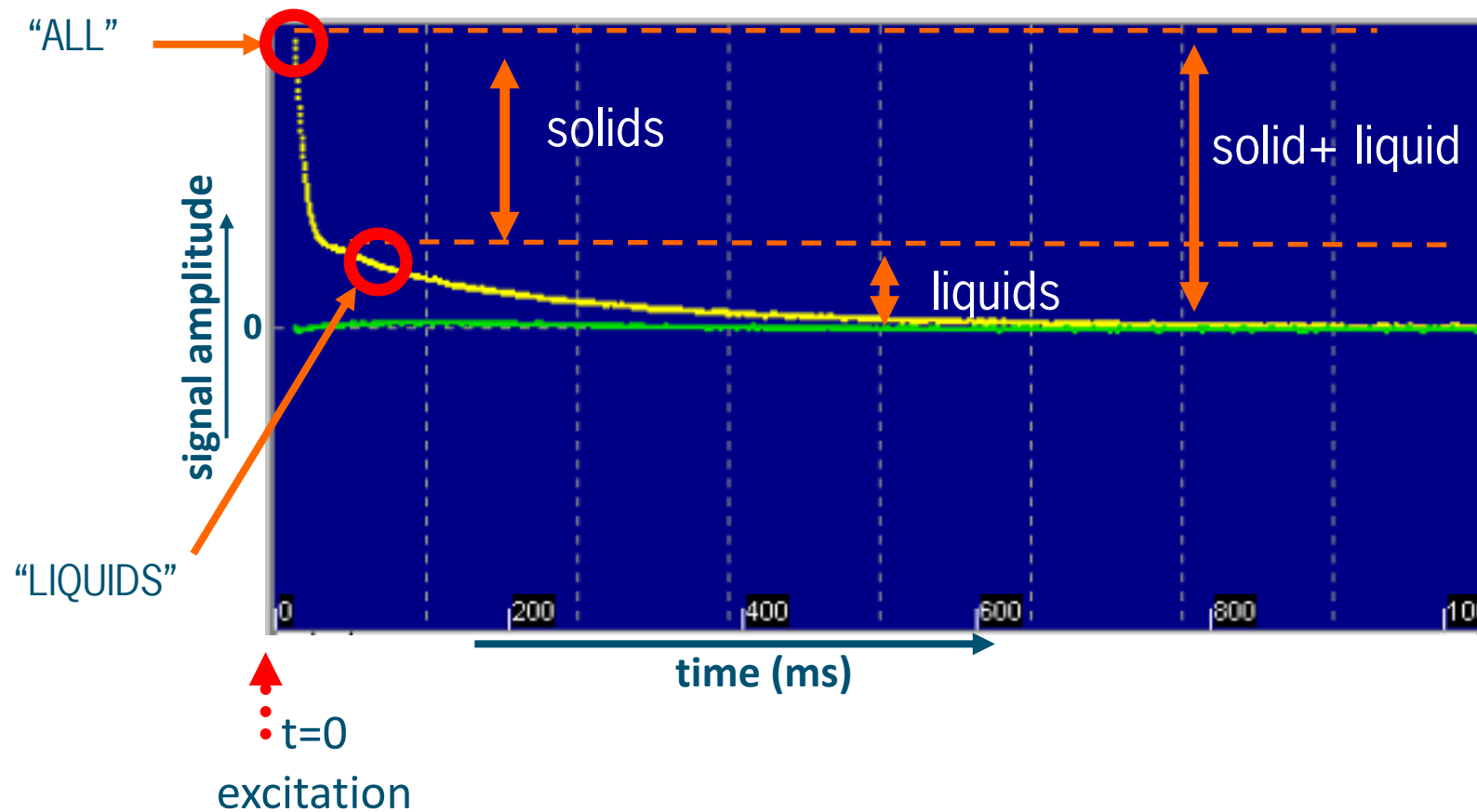
- NMR: non-spatially resolved, fast, detailed information, relatively cheap.

Problem: tissue heterogeneity

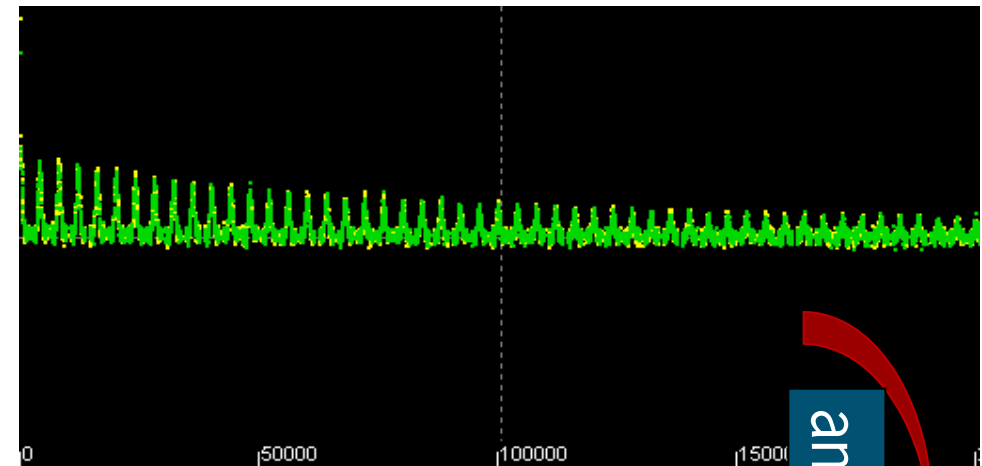
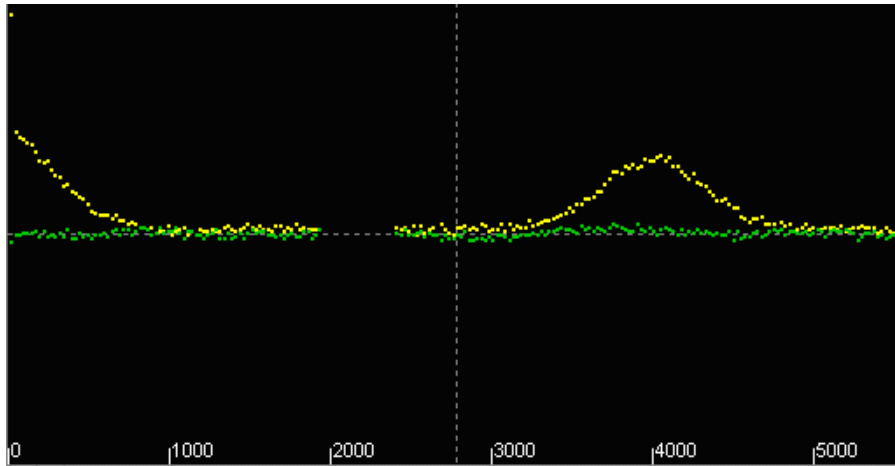
- MRI: spatially resolved, longer measurement times, less detailed information on cell level.

Problem: expensive dedicated equipment. But

The (TD-)NMR signal

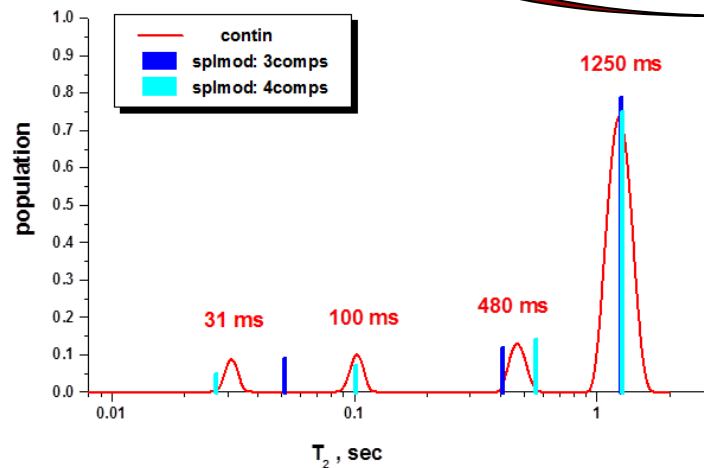


TD NMR: dynamics of water ^1H 's



One \rightarrow multi echoes

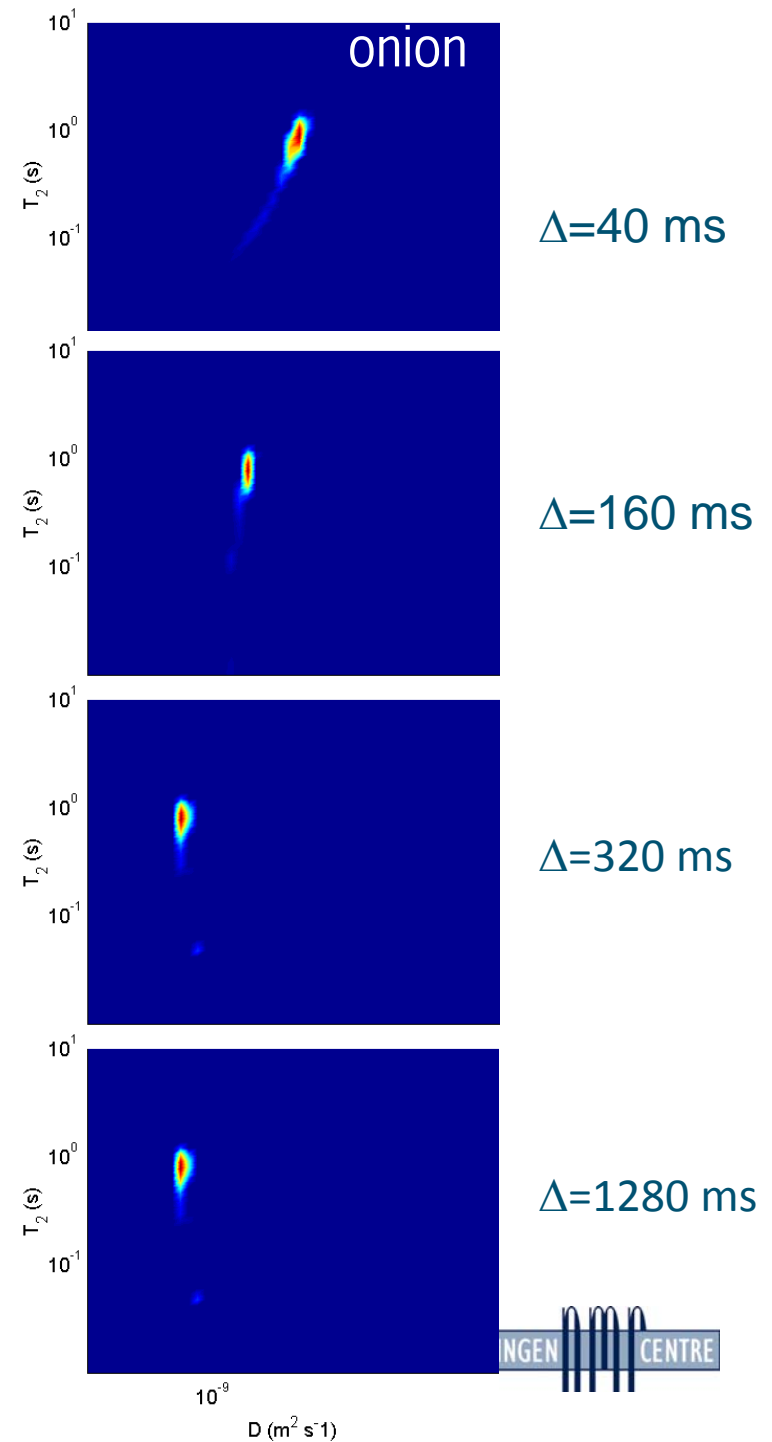
analysis



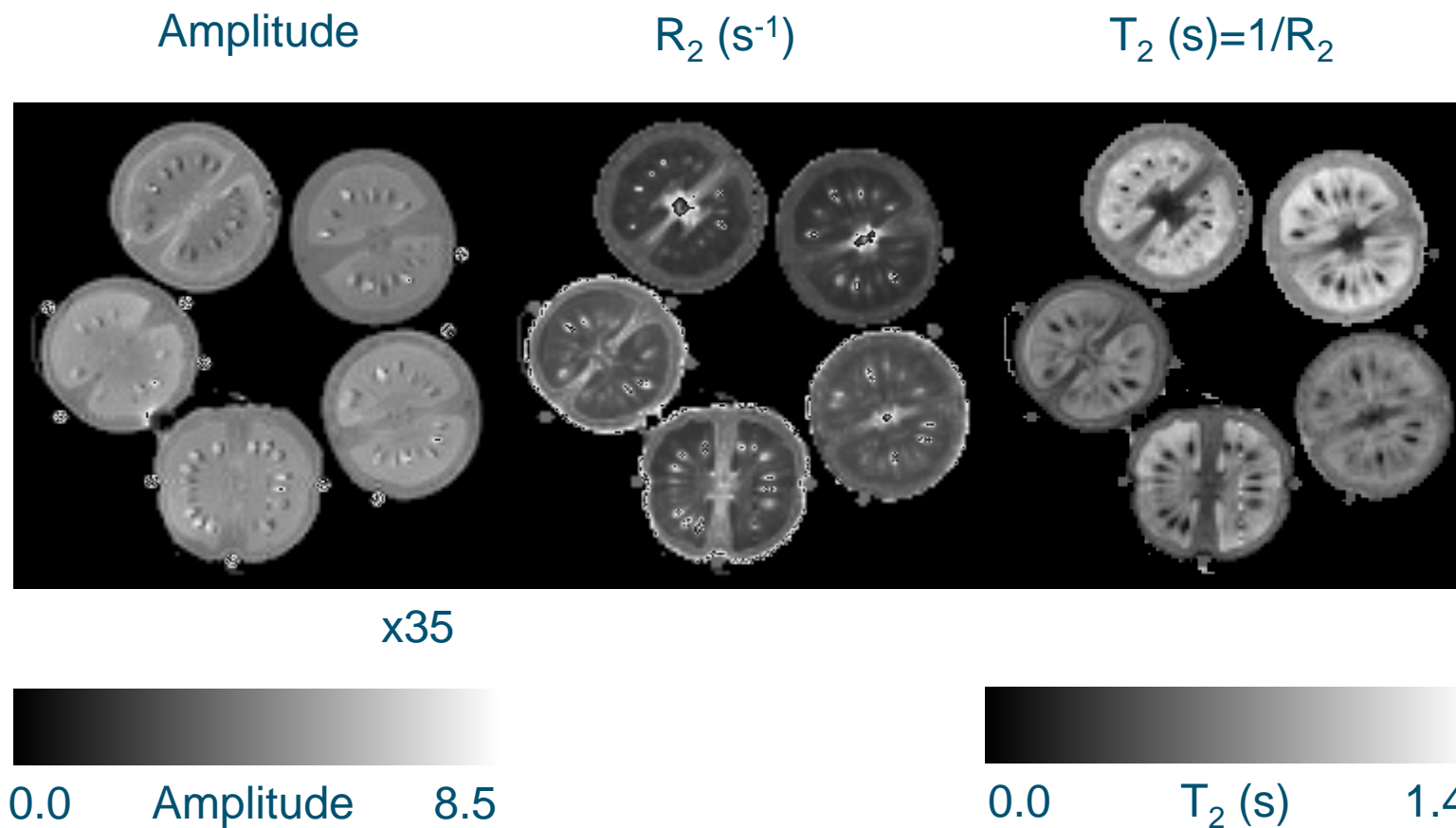
Sum of exponentials

T_2 distribution (CONTIN)

More details: use diffusion of water molecules to characterize free diffusion path length, combine with T2

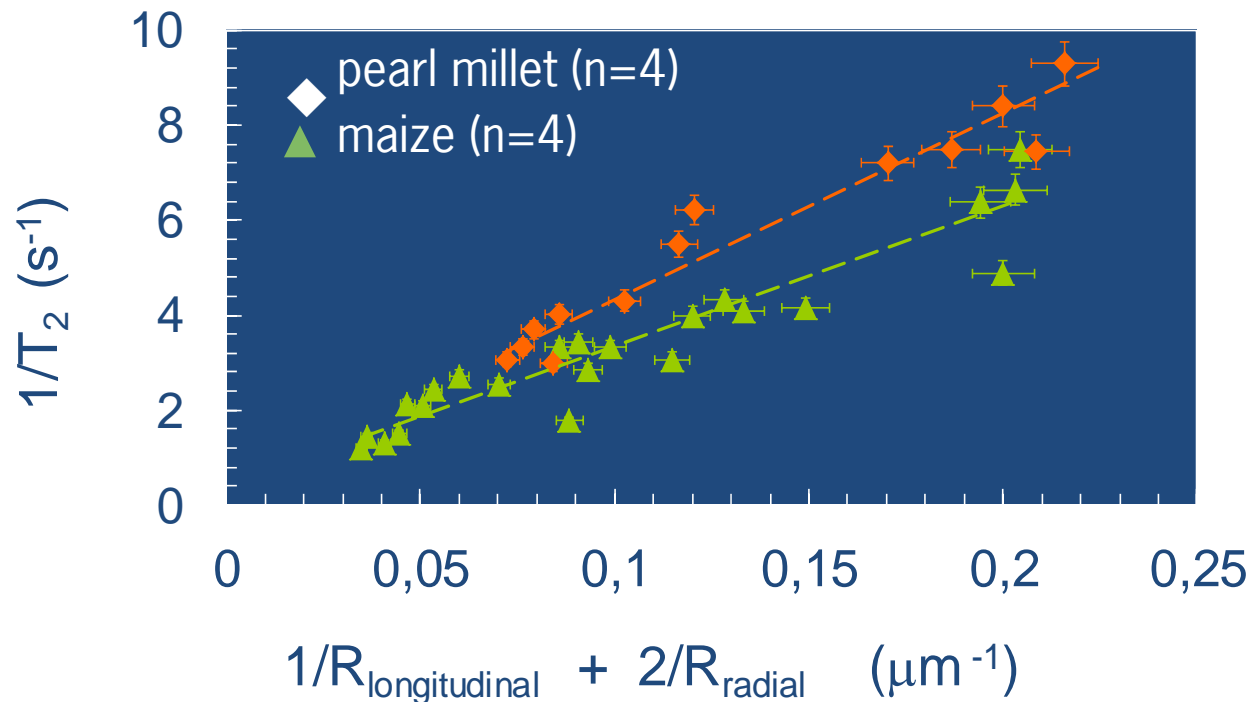


Effect of membrane permeability on T2



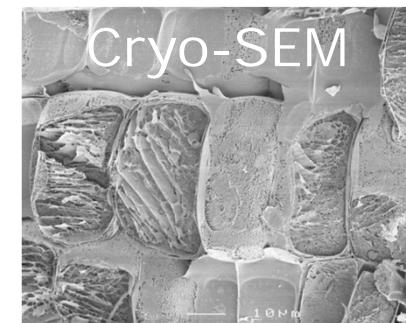
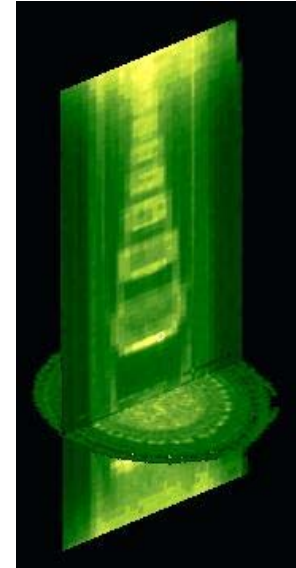
Longest T2, cell (vacuole) size and membrane permeability (H)

$$\blacksquare \quad 1/T_{2,\text{obs}} = H(1/R_x + 1/R_y + 1/R_z) + 1/T_{2,\text{bulk}}$$

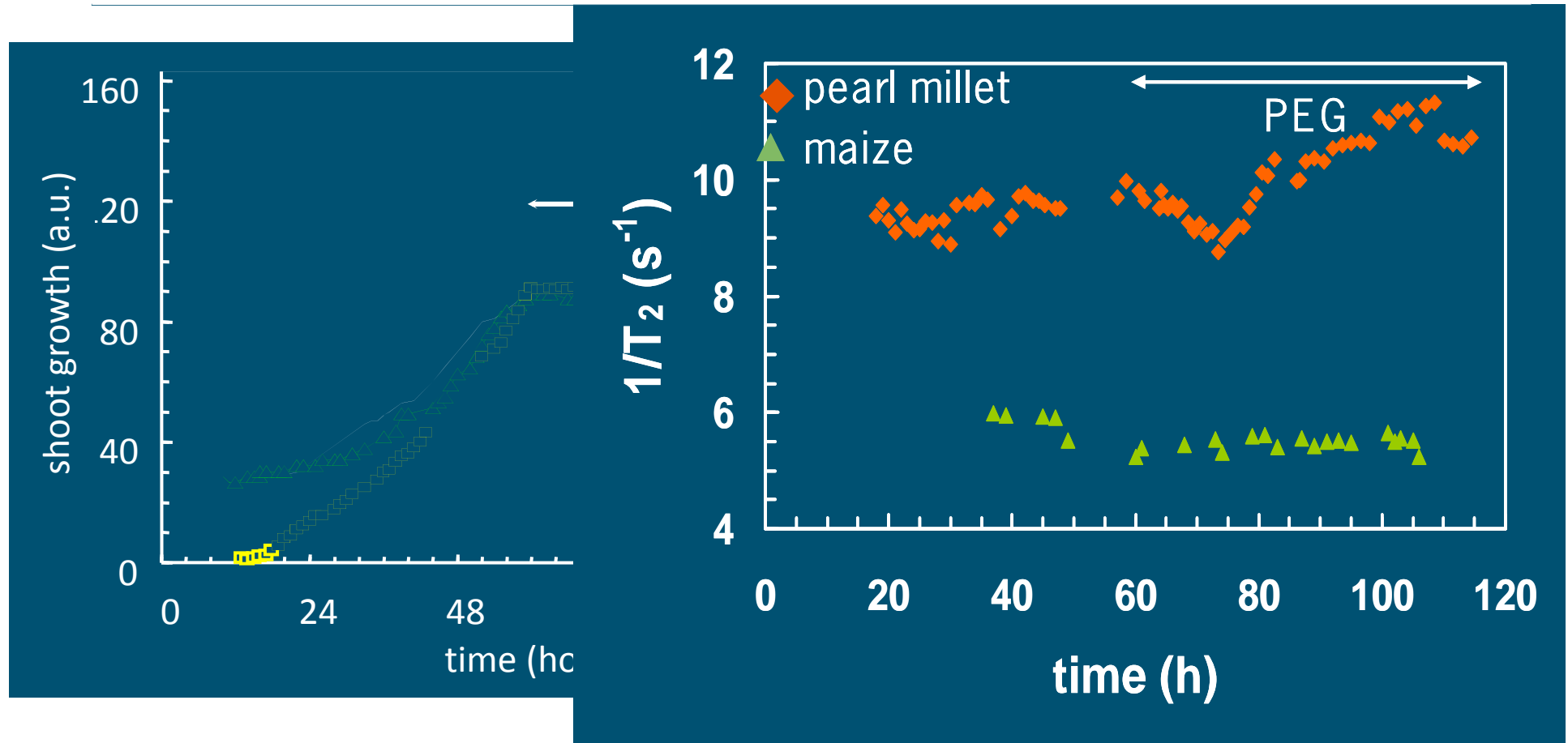


L. van der Weerd *et al.*, Plant, Cell and Environment 2002

MRI



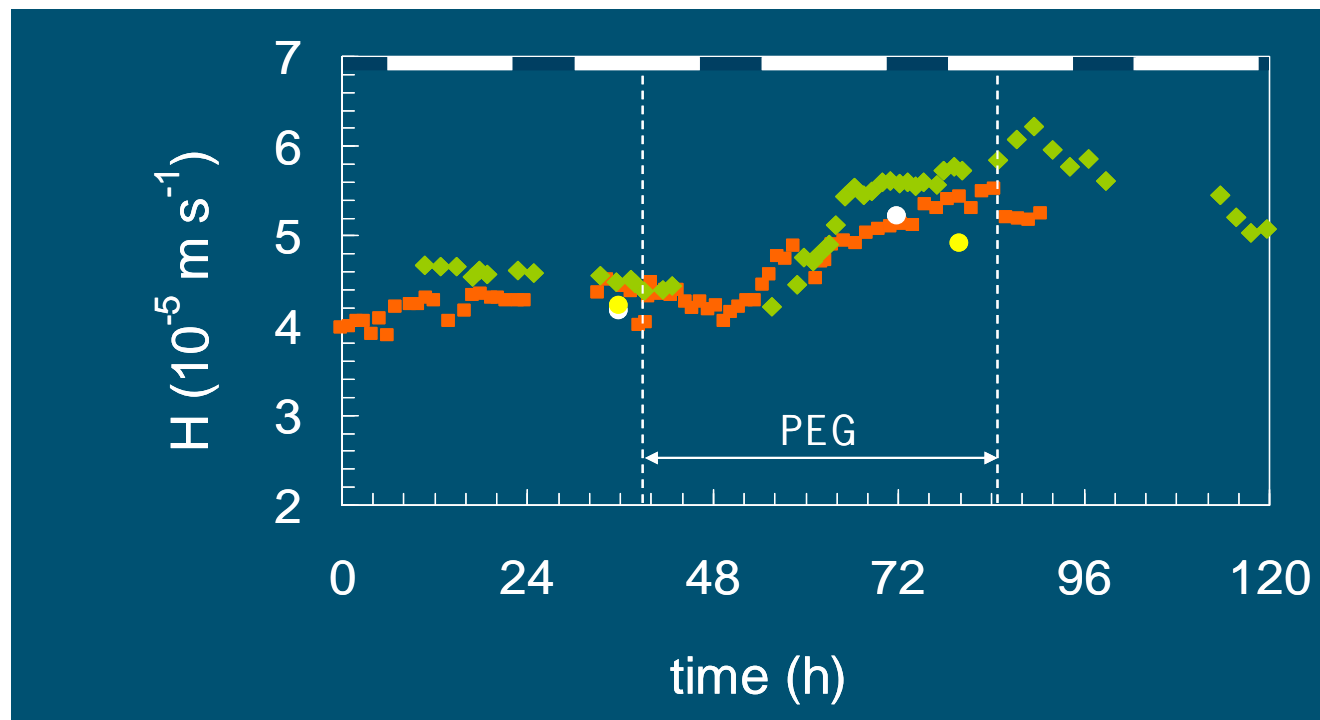
Stem (apex) growth and T_2 during osmotic stress: maize and millet



van der Weerd *et al.*, Plant, Cell and Environment 2002

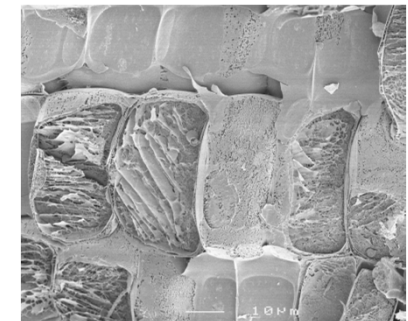
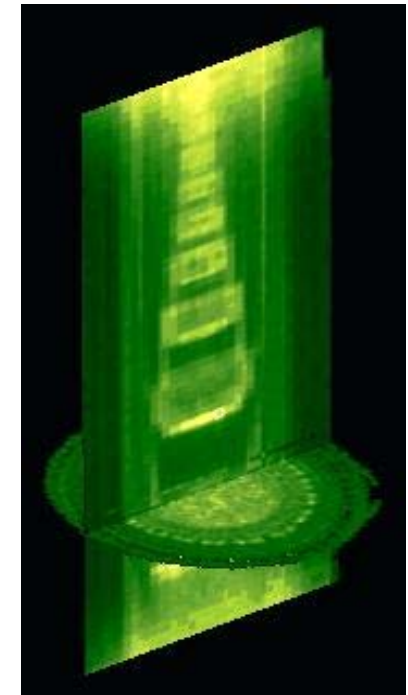
Tonoplast membrane permeability in millet during osmotic stress:

$$\blacksquare \frac{1}{T_{2,obs}} = H (S/V) + \frac{1}{T_{2,bulk}}$$



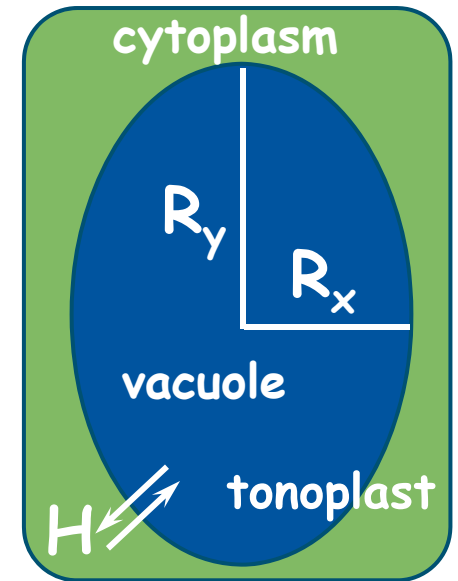
Increase in H may facilitate water redistribution between tissues
aquaporins?

van der Weerd *et al.*, Plant, Cell and Environment 2002



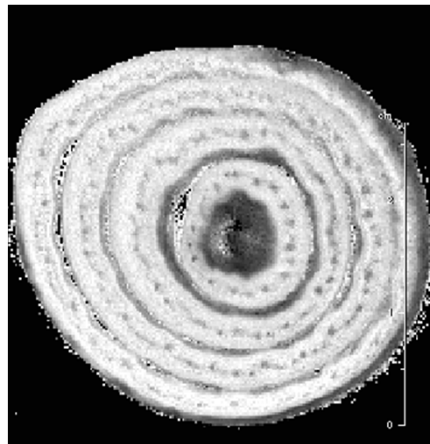
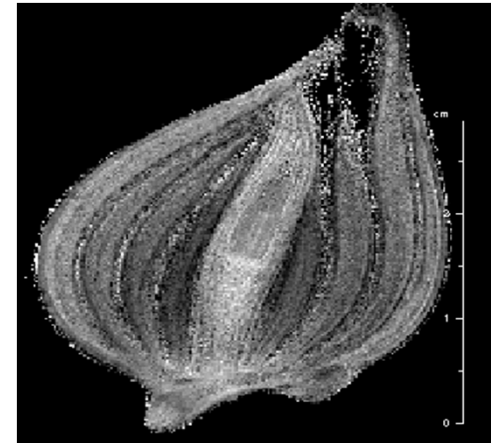
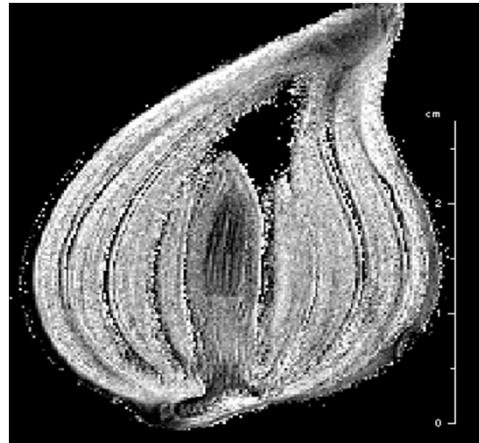
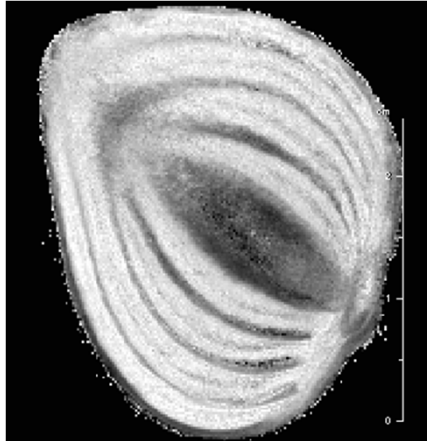
T2, diffusion and permeability

- Relaxation: $1/T_{2,obs} = H(S/V) + 1/T_{2,bulk}$
 - Can we get information about S/V?

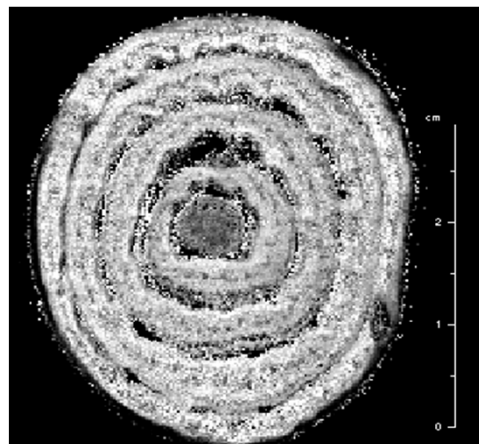


- Yes! Diffusion as a function of diffusion time:
 - Restricted due to finite compartment size
 - Short diffusion distance:
$$D_{app} (=ADC) = D_0 - S/V \cdot C \cdot D_0^{1/2} \cdot \Delta^{1/2}$$
 - Long Δ time-scale: (tissue) permeability
- Can we become independent of S/V? Yes: T1/T2

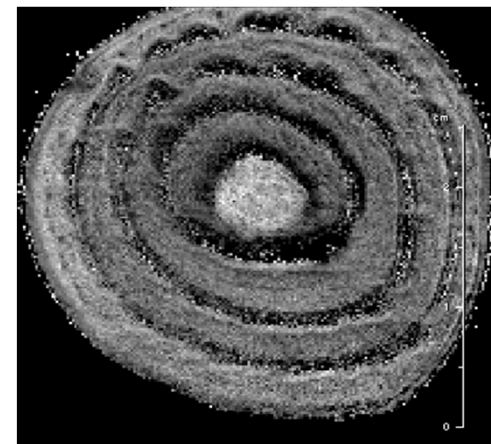
Conversion of starch: interaction between mobile and solid protons



T = 0



8 weeks



8 weeks

Transport traits: especially appealing

- Xylem: Simple proportionality (modified Darcy's law) to compare (woody) plants by xylem specific conductivity, K_s (length normalized) is rate of sap flow per cross-sectional area at a given pressure gradient:

$$K_s/(LA/XA) \propto VPD.g_s.Ht/(\Psi_s - \Psi_L)$$

- Phloem: transport between source (leaves) and sinks (roots, fruits, growing tissue,...). Can it limit optimal photosynthesis?

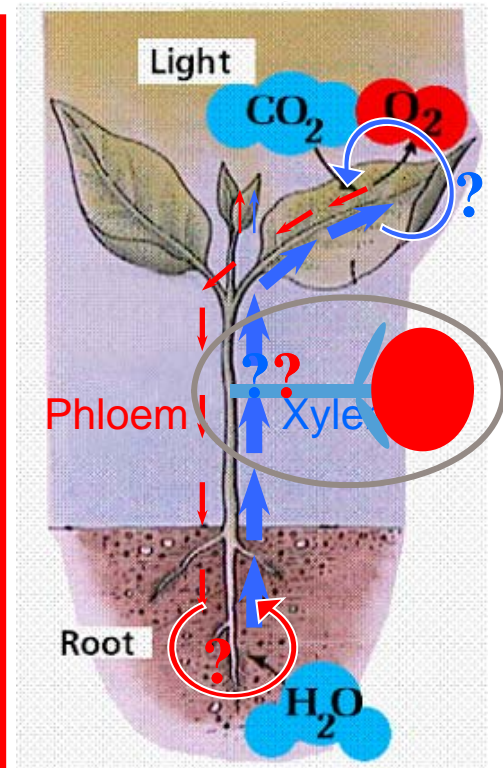
Loading from sucrose/starch.

Can we benefit from optimized unloading?

Integrated model: photosynthesis, water and sugar

Xylem and phloem sap flows are difficult to study because of their extreme sensitivity to invasive experimentation. Consequently little is known regarding the dynamics of sap flow and conductivity in an intact plant. If and when xylem transport and phloem can be limiting for optimal photosynthetic activity, CO₂ uptake and plant performance in greenhouse conditions are not yet resolved.

$$\begin{aligned}\Psi_{\text{roots}}^X &= k_{\text{soil}} \Psi_{\text{soil}}^X \\ C_{\text{crown}}^S = C_0 &\Rightarrow L = V_{\text{max},L} \frac{C_0}{K_{M,L} + C_0} \quad (\text{assumption (vi)}) \\ C_{\text{roots}}^S = C_0 &\Rightarrow M_{\text{roots}}^S = \frac{W_{\text{roots}}^S}{P_w} C_0 \quad (\text{assumption (vi)})\end{aligned}$$



(mass balance sugar)

$$C_{\text{crown}}^S = C_0 \Rightarrow M_{\text{crown}}^S = \frac{W_{\text{crown}}^S}{P_w} C_0 \quad (\text{assumption (vi)})$$

$$-f_{\text{crown}}^{X \rightarrow S} = f_{\text{crown}}^{X \rightarrow Pc} - f_{\text{crown}}^{Pc}$$

$$\Rightarrow \Psi_{\text{crown}}^S = P_{\text{crown}}^X - R_{\text{crown}}^{X \rightarrow S} \left(\frac{P_{\text{crown}}^X - \Psi_{\text{crown}}^{Pc}}{R_{\text{crown}}^{X \rightarrow Pc}} - \frac{P_{\text{crown}}^{Pc} - P_{\text{soil}}^{Pc}}{R_{\text{crown}}^{Pc}} \right)$$

$$\Rightarrow P_{\text{crown}}^S = \Psi_{\text{crown}}^S + \frac{R}{MM_{\text{carose}}} C_0 \quad (\text{mass balance water})$$

if $(f_{\text{crown}}^{X \rightarrow S} = f_{\text{crown}}^{S \rightarrow Pc} \text{ and } f_{\text{crown}}^{X \rightarrow Pc} + f_{\text{crown}}^{S \rightarrow Pc} = f_{\text{crown}}^{Pc} \text{ and } f_{\text{crown}}^{S \rightarrow Pc} > 0)$

then "start model simulation"

else "start model simulation"



De Schepper and Steppe, J Exp Bot. 2010

WAGENINGEN UR
For quality of life

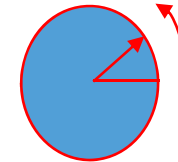
Dynamic or displacement imaging

$P(r|r', \tau)$, conditional displacement probability

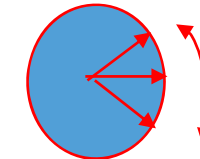
displacement Δr + gradient G

=>

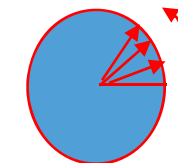
frequency or phase shift $\Delta\theta$



**diffusion, perfusion $\Delta\theta$ random,
=> no net phase shift**



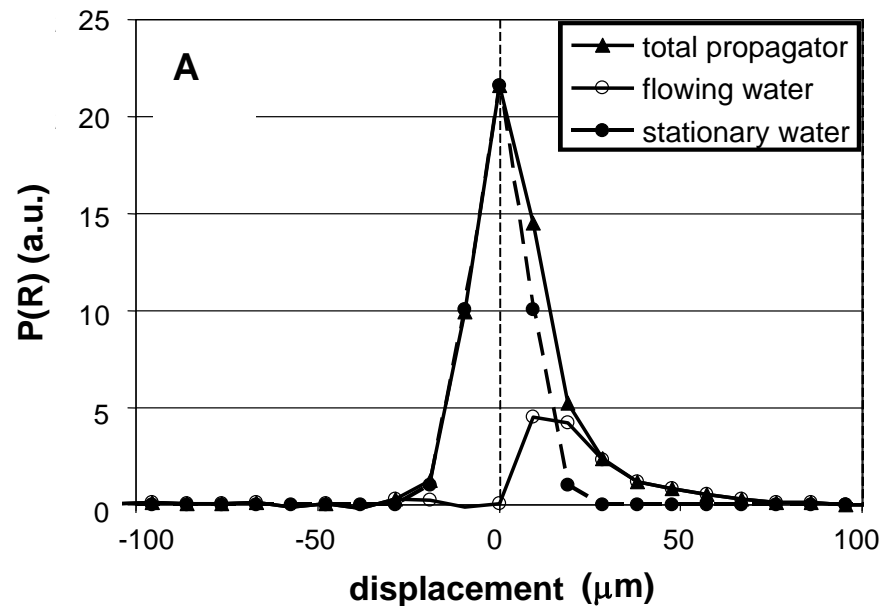
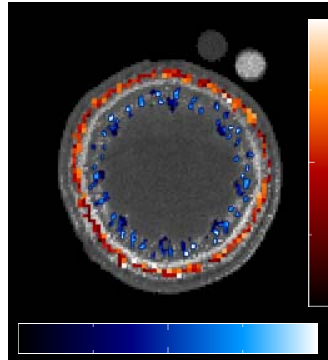
**flow: $\Delta\theta$ equal for all spins
=> net phase shift**



**detect signal as function of $G = FT$ =>
distribution of displacements Δr**

Propagator and flow characteristics

For every pixel in the image or selected regions!



Volume-flow

$$Q = \sum_{R=0}^{R=R_{\max}} P_n(R, \Delta) \cdot R \cdot \frac{A_{\text{ref}}}{\Delta}$$

Average velocity

$$v = \frac{\sum_{R=0}^{R=R_{\max}} P_n(R, \Delta) \cdot R}{\sum_{R=0}^{R=R_{\max}} P_n(R, \Delta) \cdot \Delta}$$

$$FCA = \sum_{R=0}^{R=R_{\max}} P_n(R, \Delta) \cdot A_{\text{ref}} (= Q/v)$$

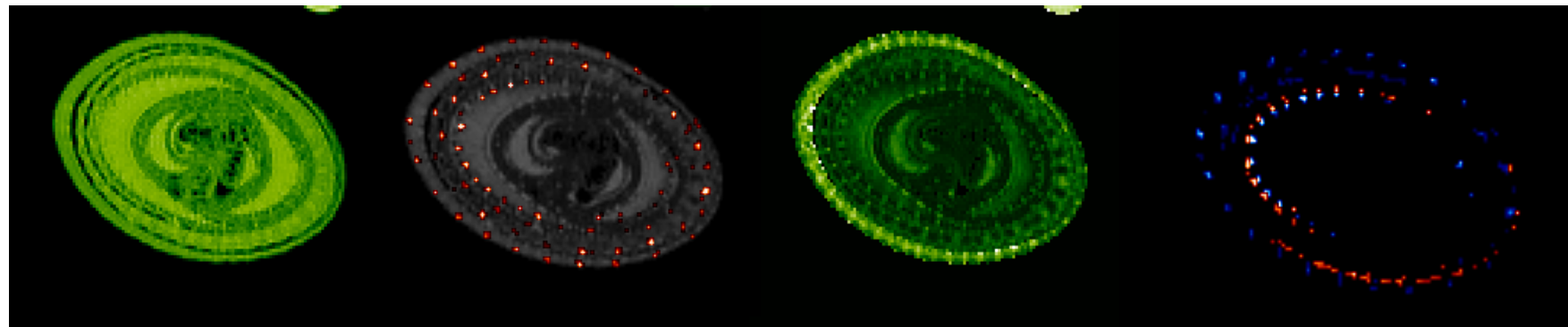
Flow: xylem and phloem in Maize

Amount of
water

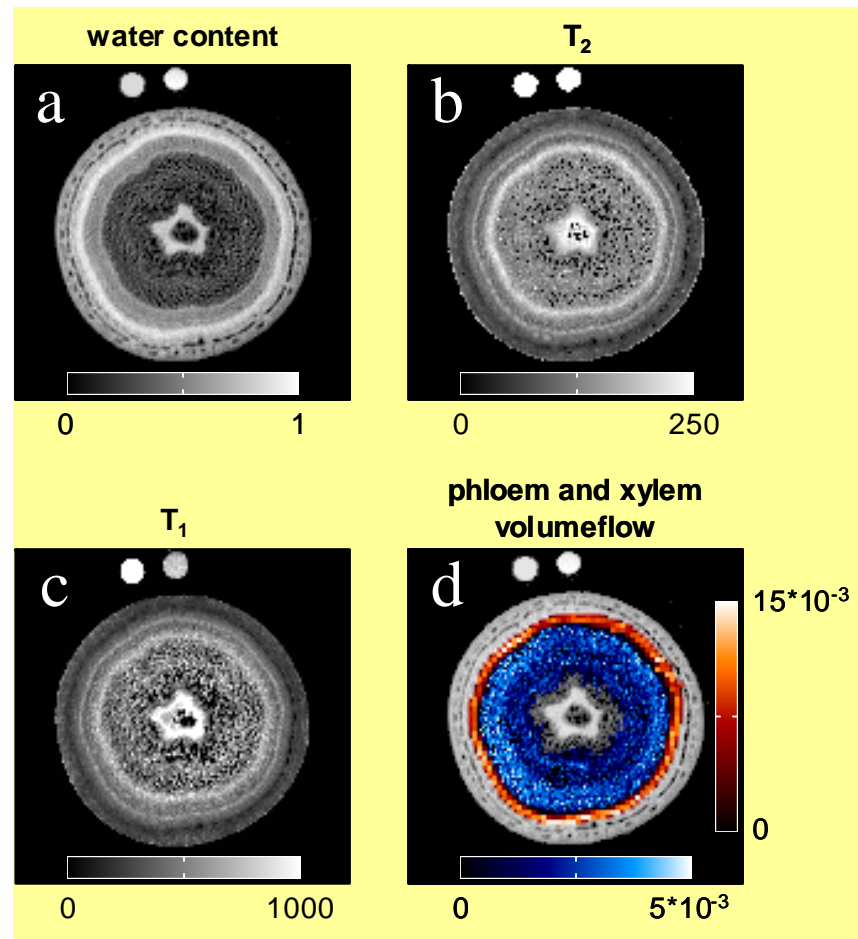
flow:
xylem (day)

T2

flow:
xylem and phloem
(night)

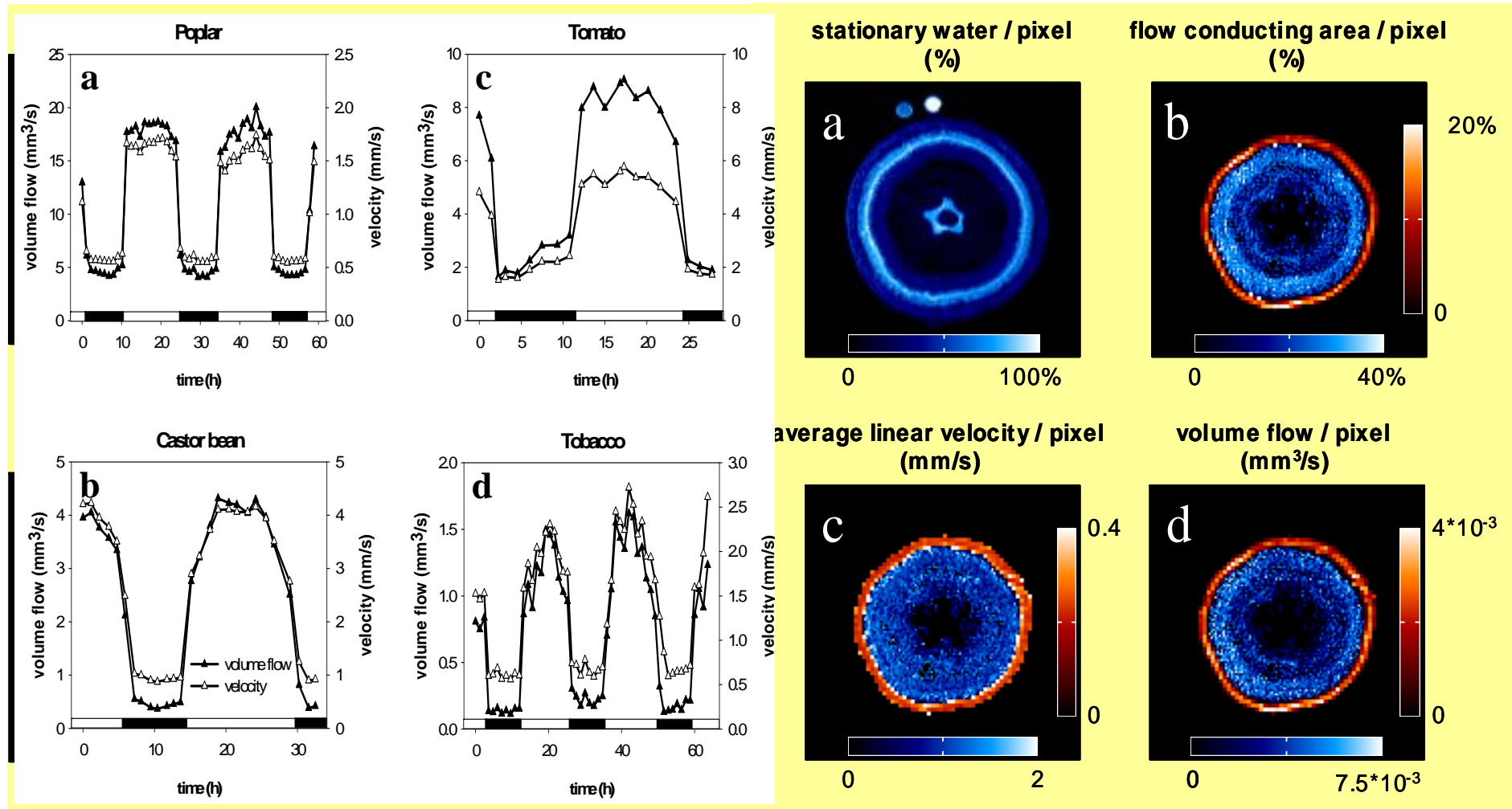


NMR/MRI: axial and radial transport and water content in storage pools



Resolution: 80 - 100 μm
Velocities:
100 $\mu\text{m/s}$ – tens of cm/s

Dynamics in xylem and phloem flow

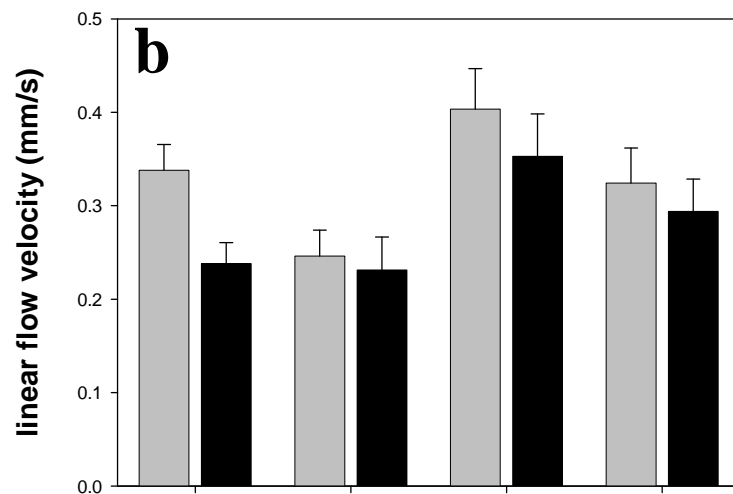
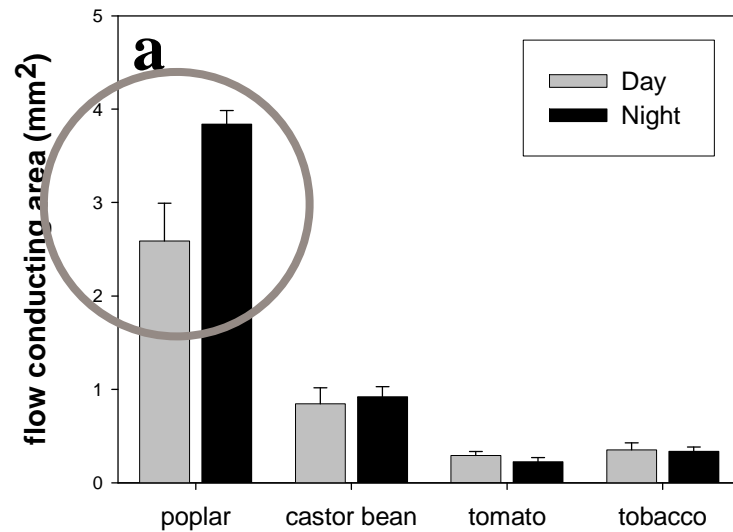


(Windt *et al*, PC&E 2006)

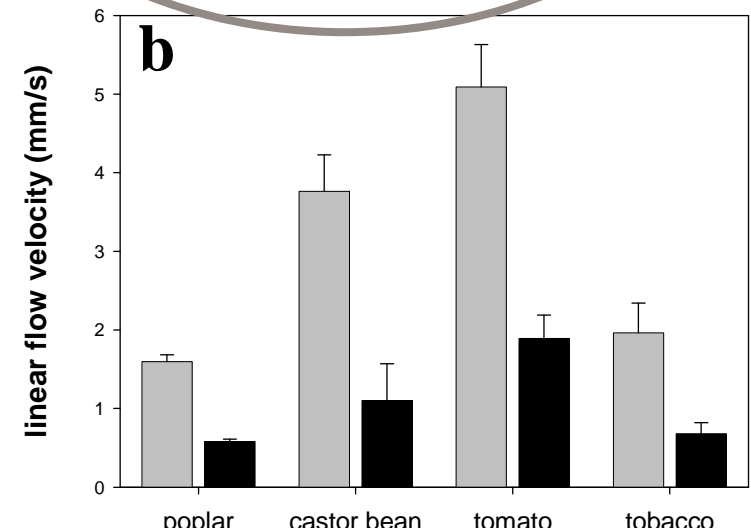
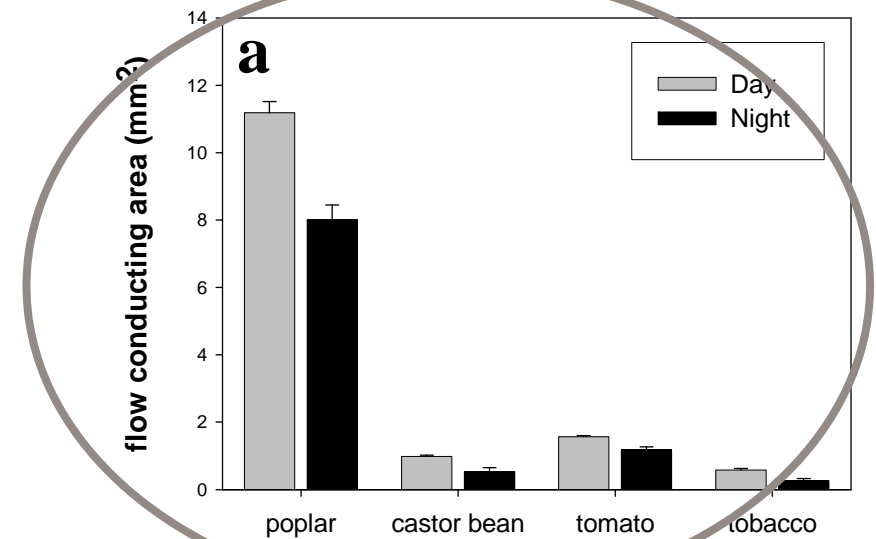
Dynamics in flow conducting area / conductance!?

(Windt et al, PC&E 2006)

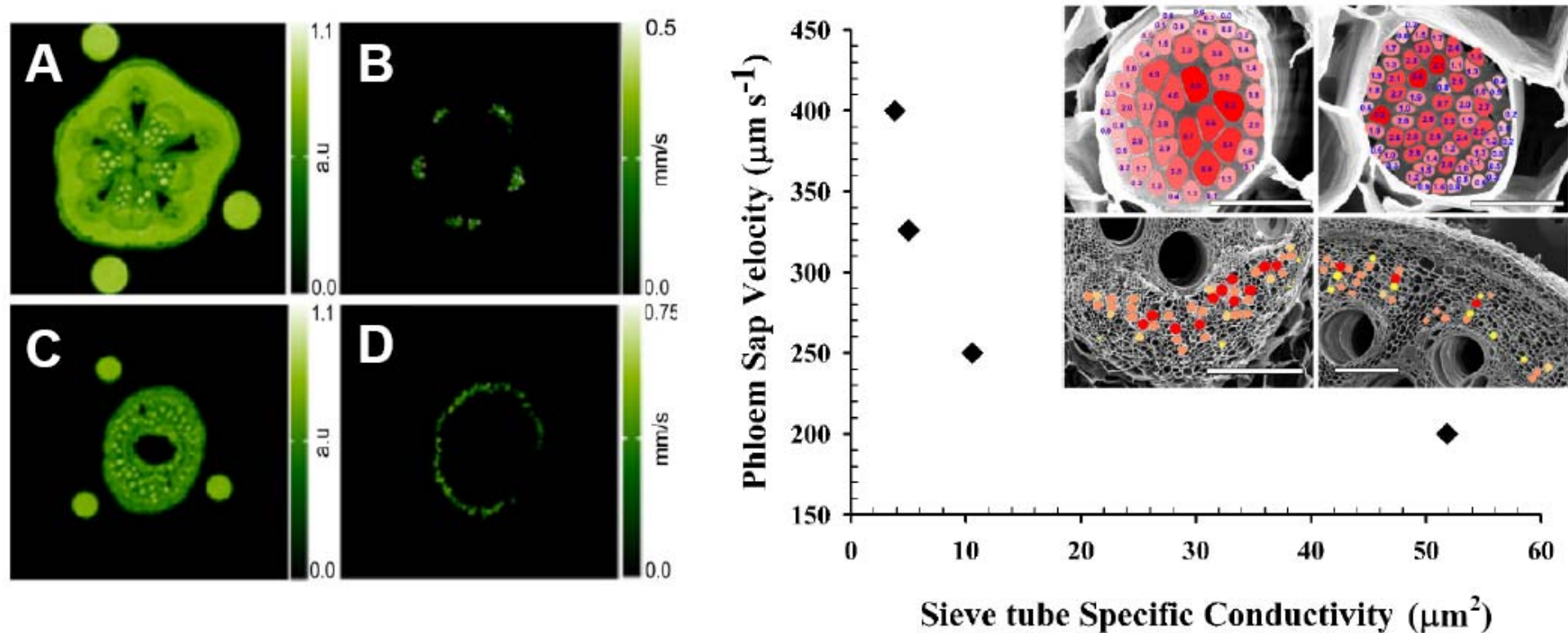
Phloem



Xylem



Phloem flow velocity can not simply be explained by conductivity and / or laminar flow?

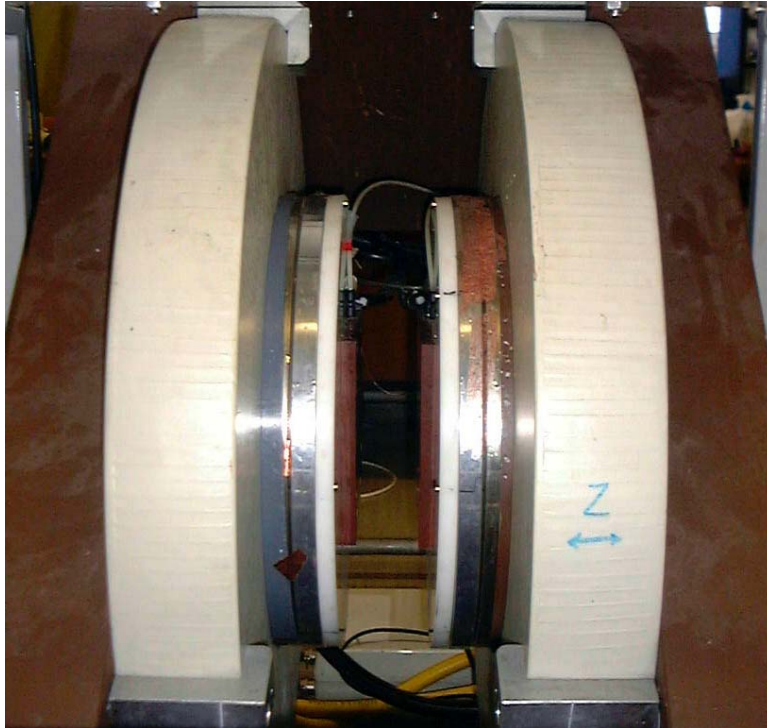


Mullendore *et al*, The Plant Cell 2010

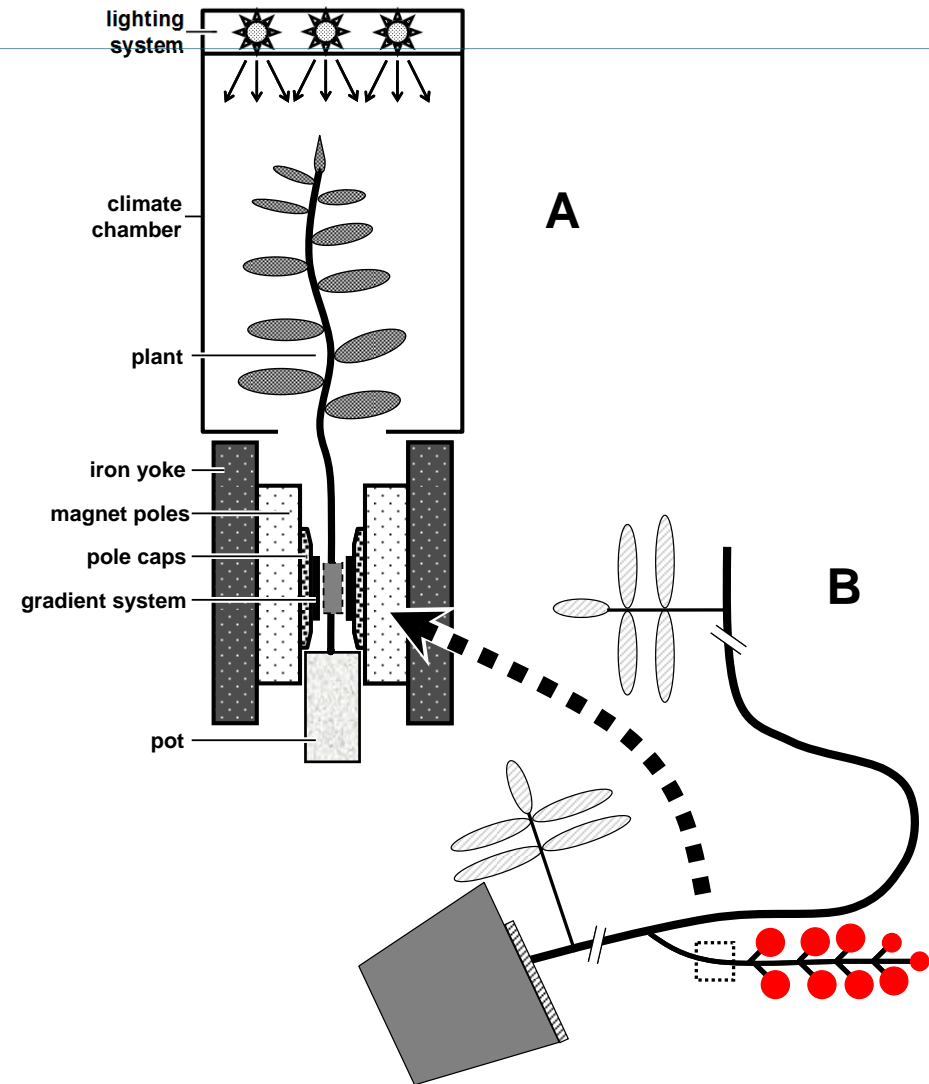
Grape: bi-directional



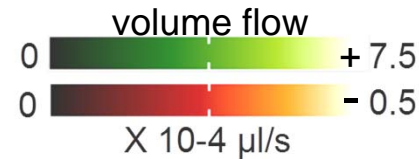
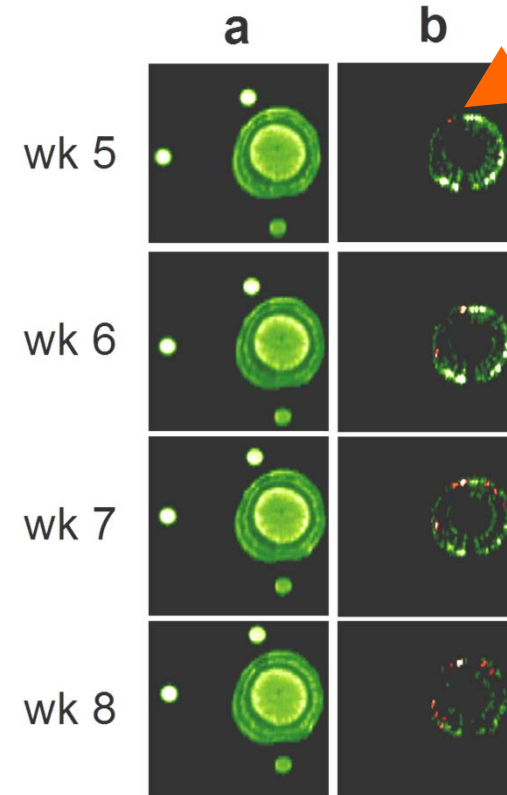
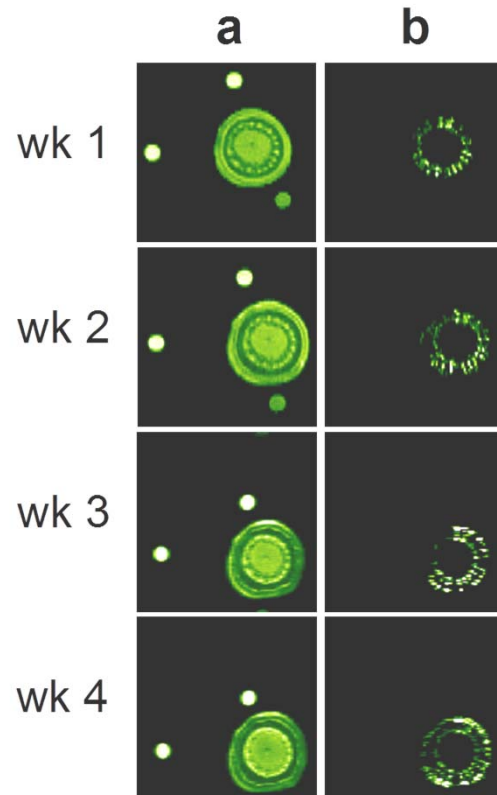
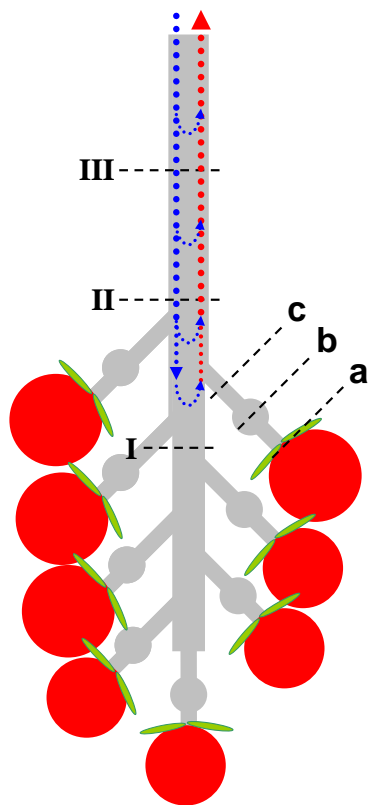
Imaging stem-truss stalk connection



electromagnet and
open gradient system



Flow in truss stalk tomato during 8 weeks of truss growth: Ratio [xylem]/[phloem] is >3:1 and not 1:9!!!



back flow

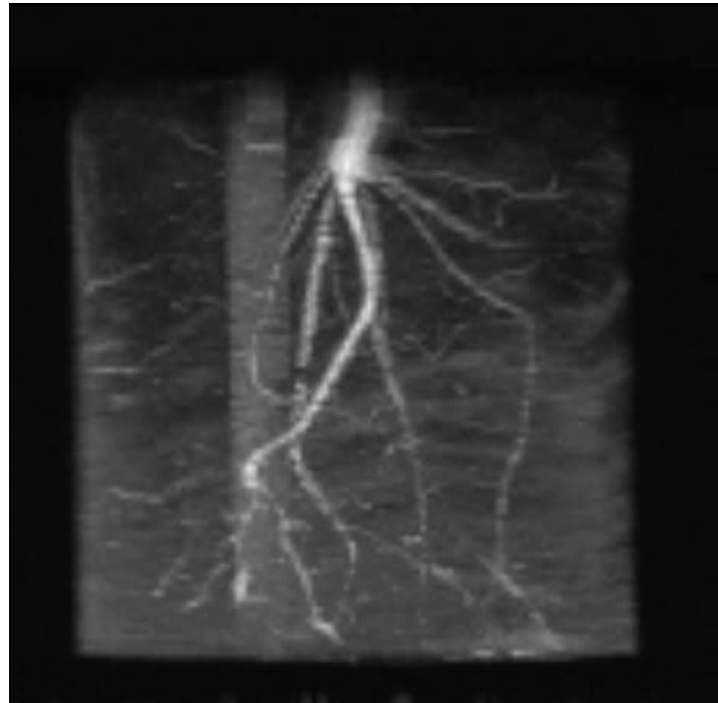
5 fruits red,
one green
remaining

last fruit red

Below ground: 3D root anatomy and water uptake



Maximum intensity projection



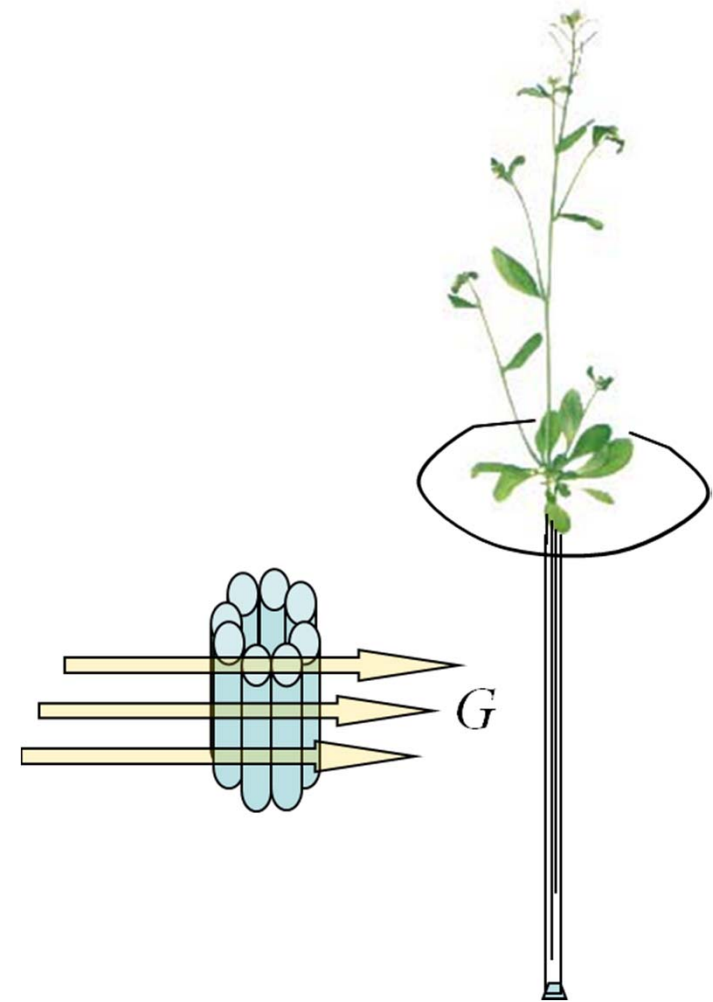
- 3T-MRI, Fast Spin Echo
- 128 x 128 x 128 pixels (FOV 10 cm -> ~ 0.8 x 0.8 mm³ per pixel)
- Total 3D experiment time 51 min 12 s

Root transport: e.g. ecotypes in relation to aquaporin expression

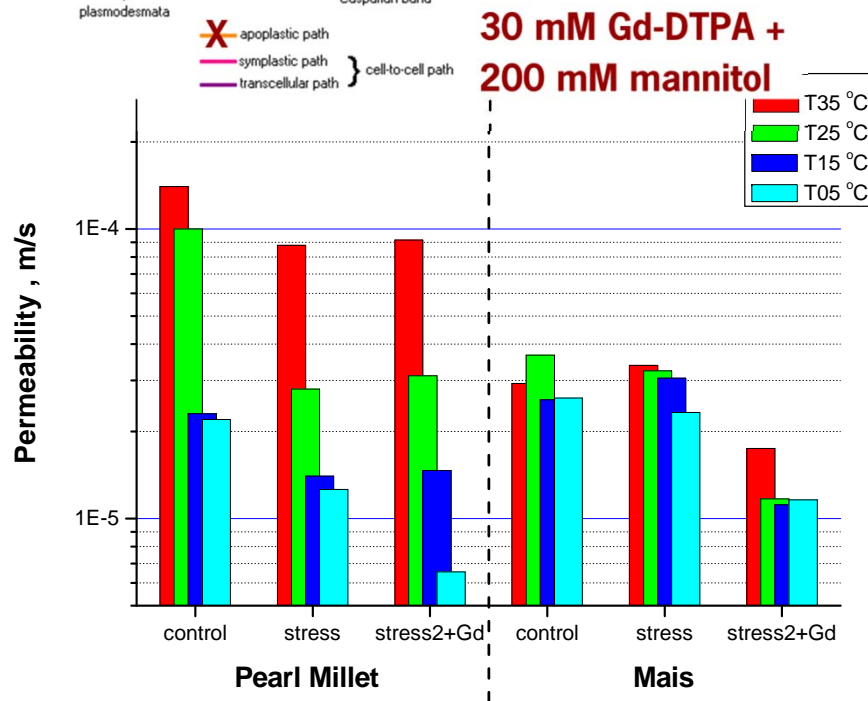
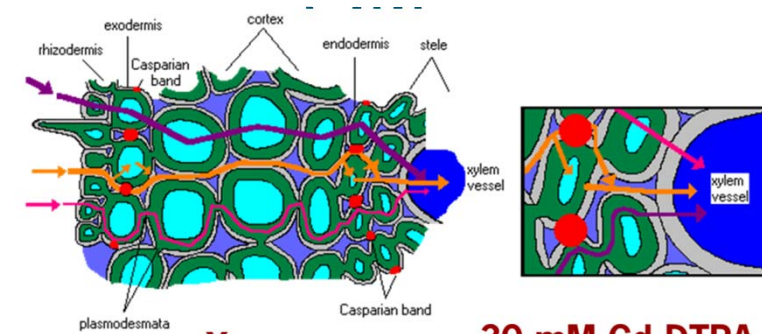
- *Arabidopsis thaliana*

ecotype: *Columbia*;

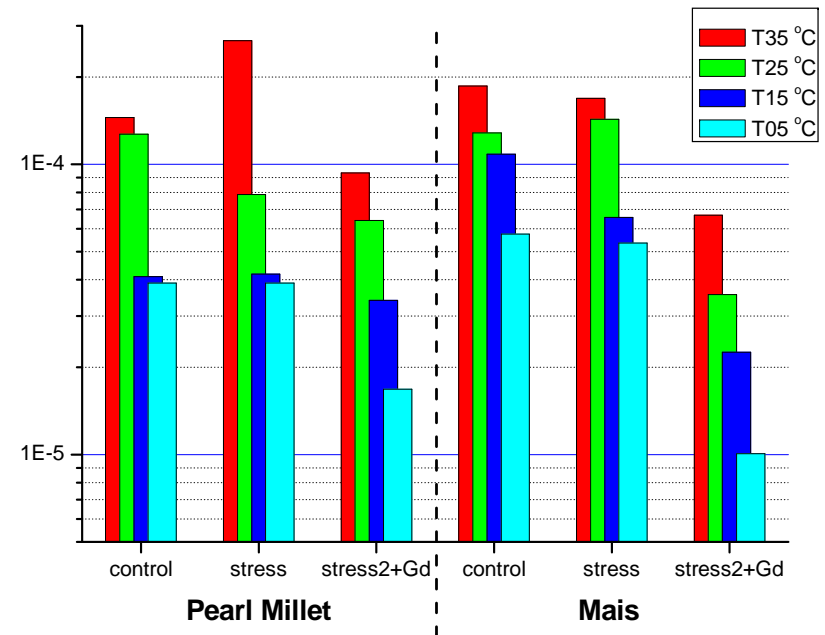
age: 4~6 weeks.



Transport in roots: pathway and tissue



Comp 2 (smaller T2 and cell size) - Millet and Mais



NMR / MRI and phenotyping

- Anatomy: leaf thickness, 3D root structure, thickness of different tissues, cell size, vessel diameter, compartments,...
- Internal parameters: water content in different tissues, transport and flow conducting area (xylem and phloem; sink-source connection), membrane or tissue permeability, starch content (leaves, beans), ...
- Static versus dynamics (growth and adaptation processes and rate)

MRI

- ❖ Lab bound, but ... starting to become (trans)portable



Dedicated Plant MRI systems and climate control: 0.7 and 3T



lights

climate chamber

plant

gradient probe

electromagnet

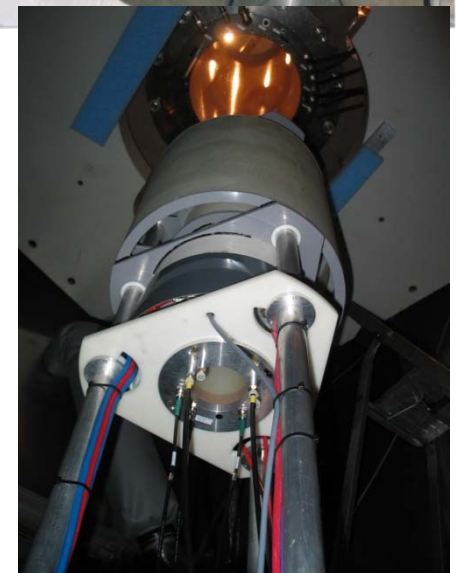
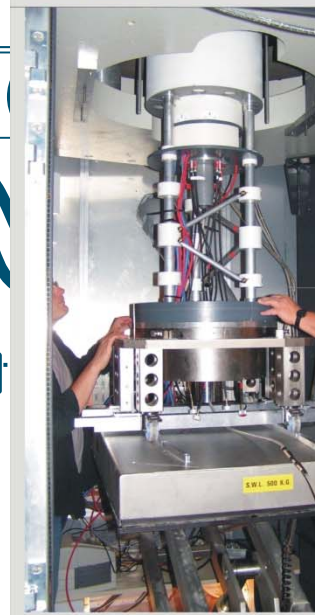
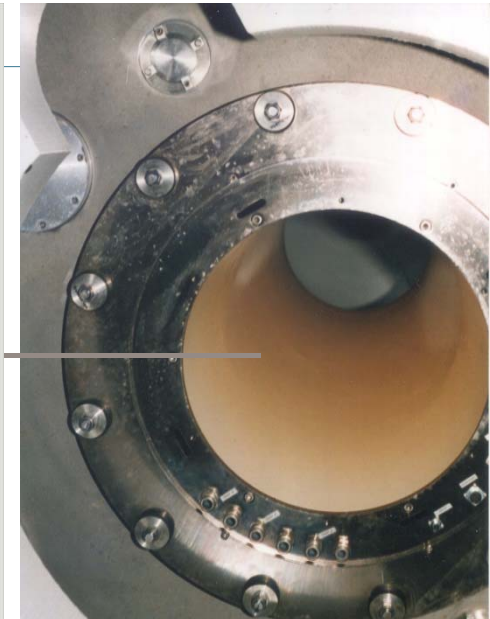
Growth

medium

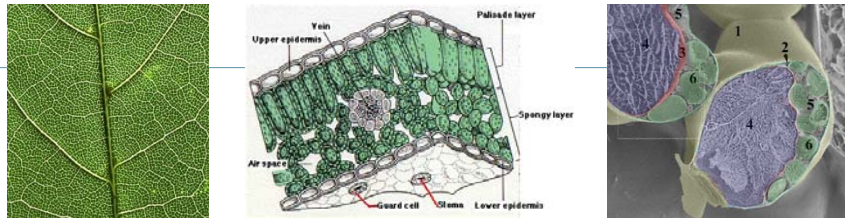
Spiral

cooler

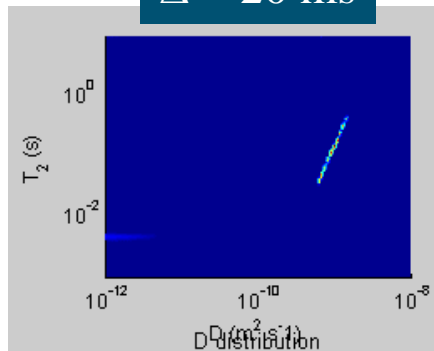
Balance



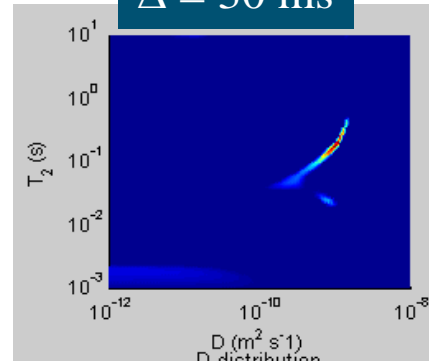
Chloroplasts, photosynthesis and single-sided portable NMR



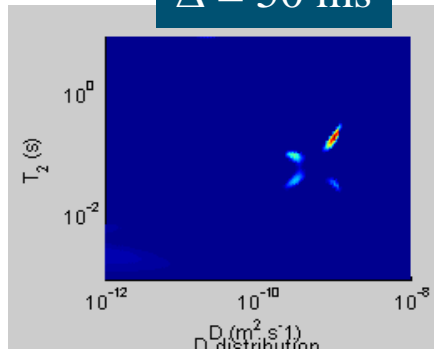
$\Delta = 20$ ms



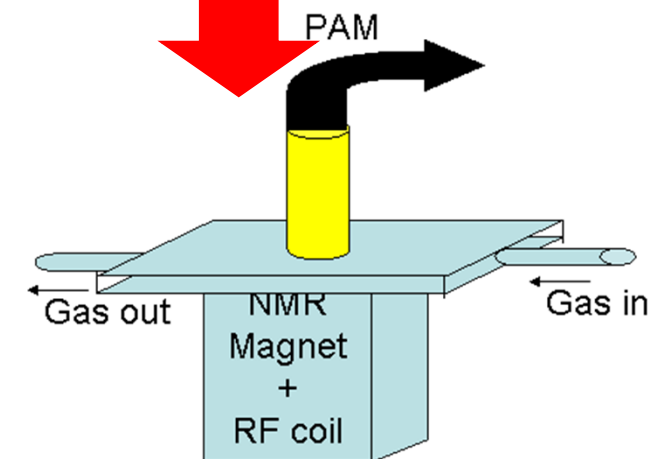
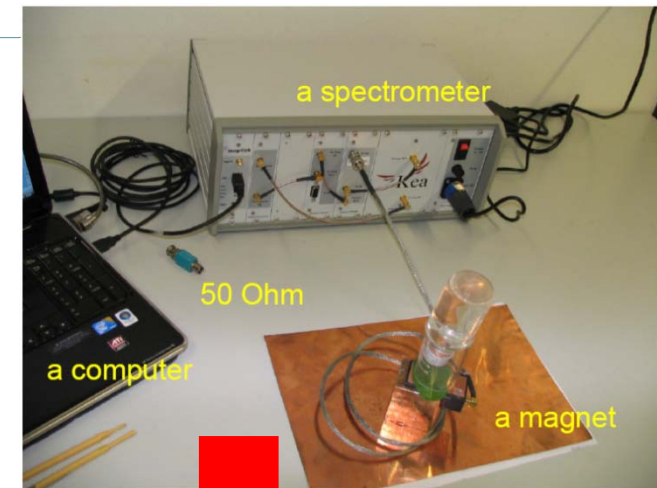
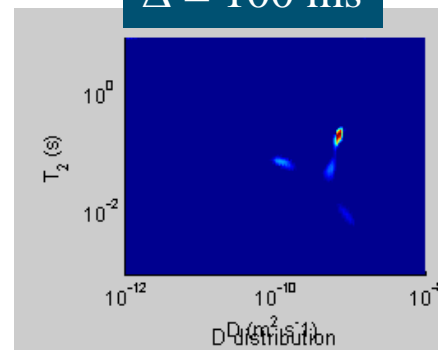
$\Delta = 30$ ms



$\Delta = 50$ ms



$\Delta = 100$ ms



Rascher et al Func Plant Biol 2011

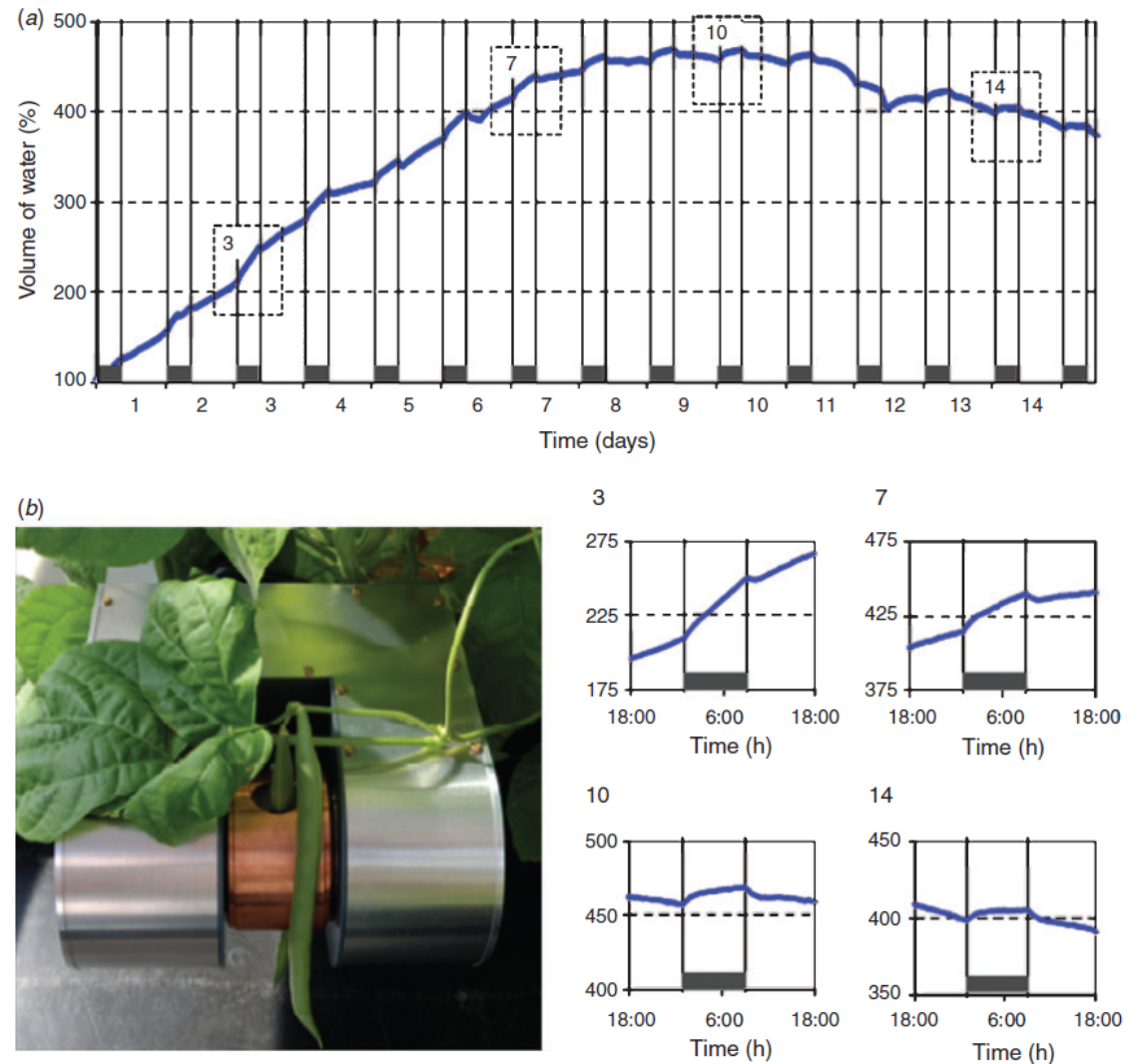


Fig. 4. (a) Quantifying shoot transport by portable nuclear magnetic resonance (NMR). Growth of a bean pod during a period of 2 weeks. Shown is the relative change in the volume of water in a 15 mm section in the middle of the bean pod, as placed within the coil of a custom built portable NMR scanner. The bean pod was 7 days old at the start of the measurement. Four day-night-day transitions are shown in detail, at night number 3, 7, 10 and 14 (indicated with dashed lines in the main graph). The night periods are indicated with dark grey bars. (b) Image of the C-shaped NMR magnet is shown, fitted with a sample holder containing a bean pod. During pod growth the environmental conditions were as follows (climate chamber): day 23°C, RH 65%, 300 μ E, 16 h; night 23°C, 65% RH, 8 h. NMR hardware and settings: custom built portable C-shaped magnet, 10.35 MHz, air gap 30 mm, temperature kept constant at 25°C \pm 0.1°C by means of an electronically controlled heater. The magnet was insulated to protect it against sudden temperature changes. For the RF coil a solenoid with an inner diameter of 15 mm and 13 windings was used. Magnet and coil were used in conjunction with a Magritek Kea 2 spectrometer with integrated RF amplifier. For each cycle a CPMG type measurement was run, with a repetition time of 7.5 s, 3000 echoes, eight complex points per echo, 32 averages and a spectral width of 100 KHz; total scan time for each point was 4 min.

Mobile MRI: Kimura *et al*, 2011

053704-2 Kimura *et al*.

Rev. Sci. Instrum. **82**, 053704 (2011)

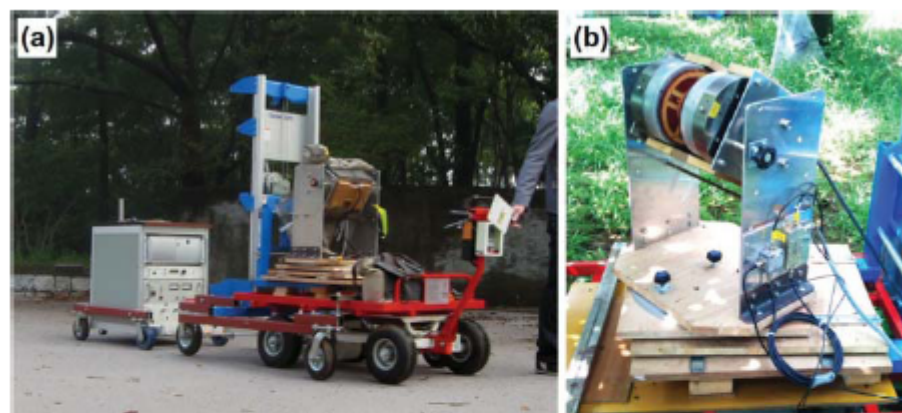


FIG. 1. (Color online) (a) Overview of the electrically motorized mobile MRI system using a permanent magnet. (b) Close view of the permanent magnet with two rotation axes and two horizontally sliding tables. The rotation axes are vertical and horizontal axes.

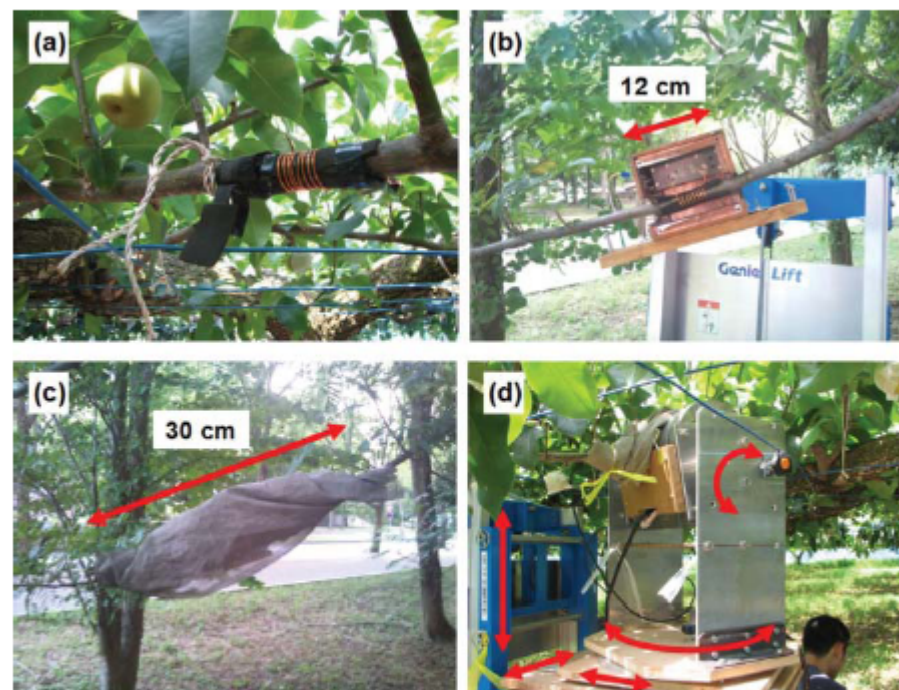
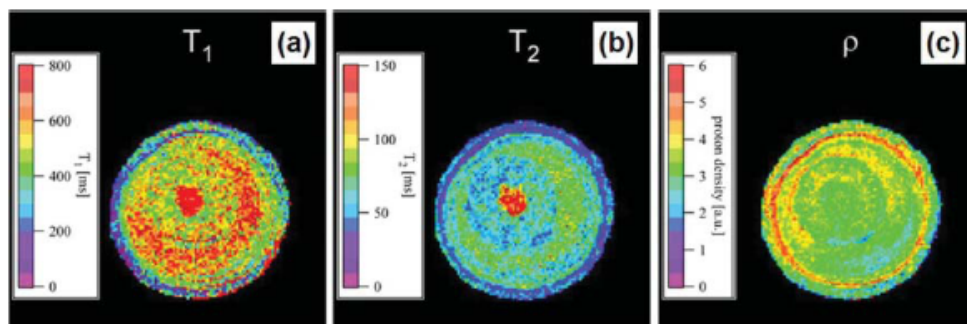
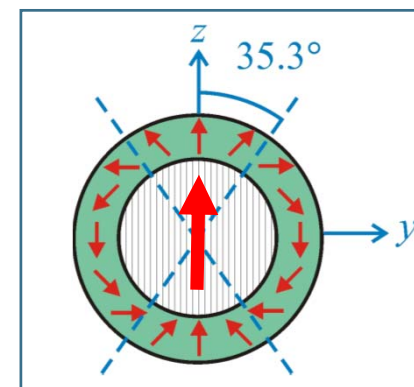
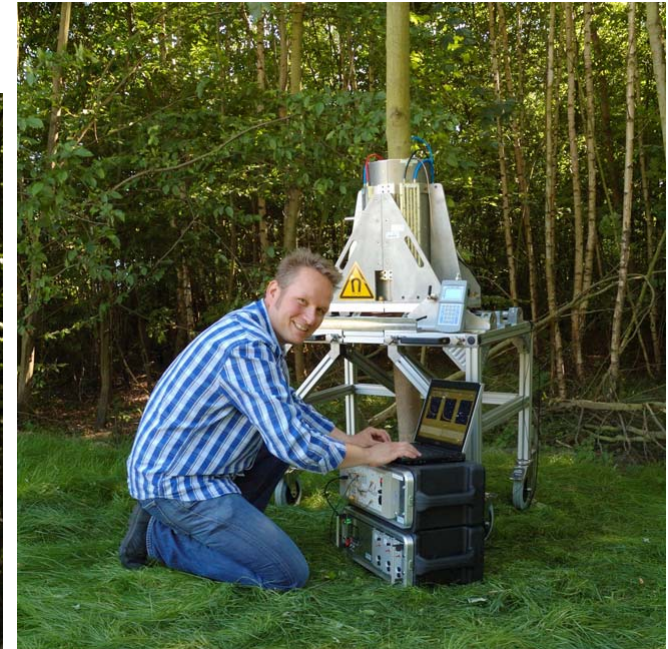
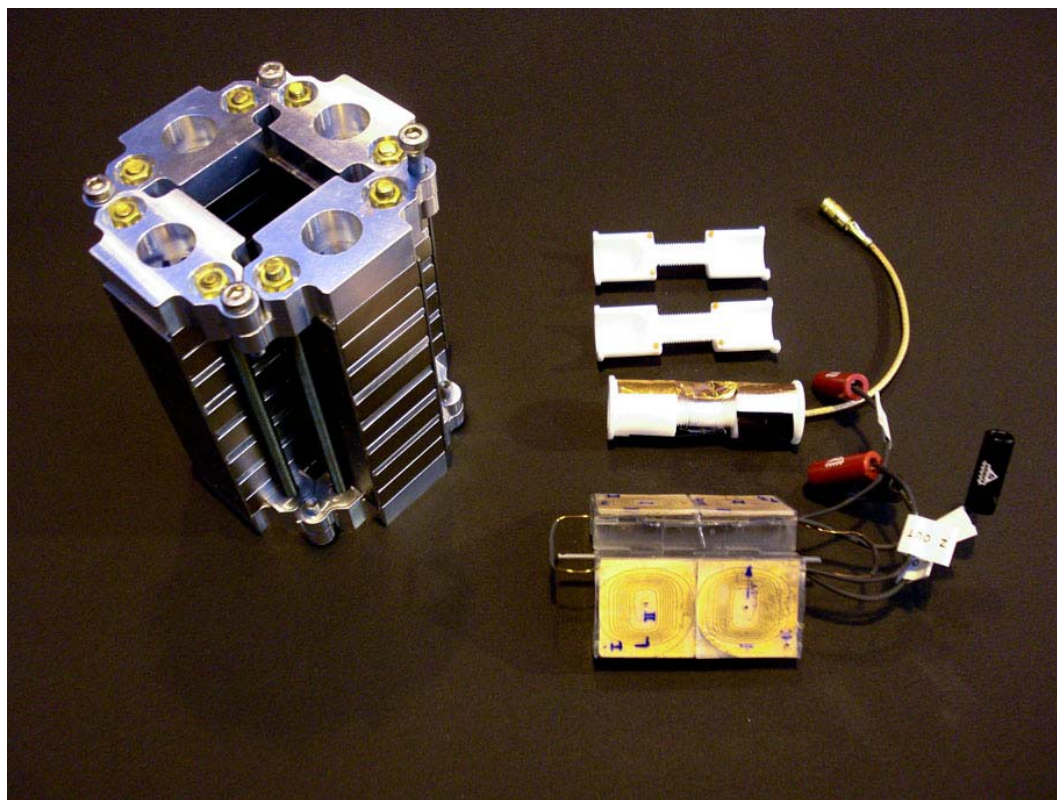


FIG. 2. (Color online) (a) RF coil wound around a branch of a pear tree. (b) Open view of the RF probe. (c) Electromagnetically shielded RF probe with a shielding cloth wound over the RF probe. (d) Setup of the RF probe in the gap of the permanent magnet. The magnet has two rotation axes and three translational mechanisms.

Halbach Tree scanner



Hinged Halbach (NMR-CUFF) and U-shape magnet



Windt *et al*, JMR 2011
Rascher *et al*, 2012



Tree Hugger

- An open access transportable 1.1 MHz ^1H MRI system for the *in situ* analysis of living trees in the forest. Access up to 21 cm. Magnet: 55 kg.

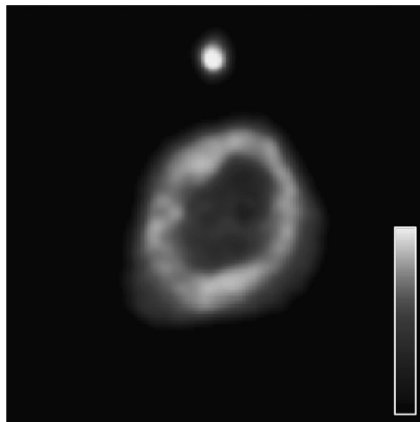


Fig. 3. Cross section image of the bird cherry tree, recorded September 2011. The white dot towards the top of the image is the reference tube of water.

Jones *et al*, JMR 218: 133-40 (2012)

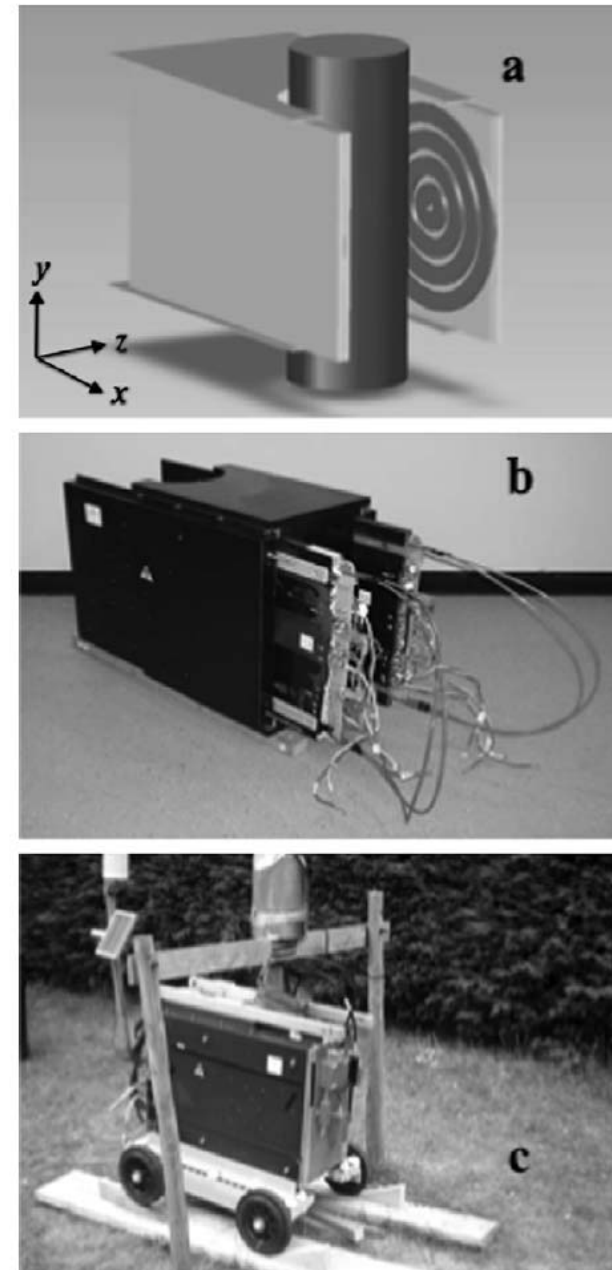


Fig. 1. a: Schematic of magnet design showing open access frame and concentric ring design (4 of 5 rings shown) of magnet with a mock tree between the poles; b: magnet as built with the gradient plates pulled half way out and c: magnet around a tree at Forest Research, Farnham, UK.

Conclusions

- Large number of parameters, some quite unique, is available by both NMR and MRI.
- The only systematic application (to my best knowledge) is MRI of 3D root anatomy (24 plants/day, can be increased to max 50 plants per day)
- There is a strong potential, but protocols and hardware have to be further developed
- This can only be realized via pilots combining NMR/MRIs and plant scientists

Acknowledgements:

Carel Windt

Natalia Homan

Frank Vergeldt

Edo Gerkema, John

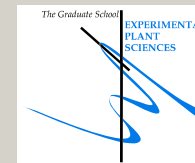
Philippi

Alena Prusova

Louise van der Weerd,

Tom Scheenen, Timur

Sibgatullin



NWO, LNV, WUR, SENTER, EU